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Synthesis, Antiglycating and Antioxidant activities of Triazolo hexahydroquinazolines

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Abstract: Some 5-aryl-triazolo[4,3-a] hexahydro quinazoline derivatives (4a-4e & 5a-5e) were synthesized and evaluated for their in vitro antioxidant and activities. compounds were found to have potent antiglycating activity than rutin in the range between 0.096 and $0.300~\mu M. The$ compounds ${\bf 4b}$ and ${\bf 5c}$ were the most active ones among them. The tested compounds had shown potent to moderate antioxidant activity by DPPH assay than BHT except **5e** (p-NO₂ phenyl derivative). Among all, hydroxyl derivatives (4a-4e) are more potent than the corresponding thiols (5a-5e).

Keywords: Hexahydroquinazolines; Triazole; Antiglycating agents; Antioxidant agents

1. Introduction

The incidence of type-2 diabetes is increasing at an alarming rate and about 1–2% of the world's population is affected with over 100 million diabetic patients worldwide. Diabetes is a multi-factorial disease with associated complications. However, exact molecular basis of these complications is not fully understood yet. Hyperglycemia is one of the contributing factors of these complications i.e. atheroma, hypertension and microangiopathy, mostly attributed to the formation of sugar-derived substances called advanced glycation end products (AGEPs) 1. A vast number of evidences suggest that AGEPs are important physio-pathogenic mediators of almost all diabetic complications 2. AGEPs are a group of molecules formed in a non-enzymatic glycation from combination of reducing sugars with free amino groups of proteins, lipids, enzymes, phospholipids and nucleic acids.

Initial product of amino group of protein with carbonyl group of glucose interaction, is a Schiff base, whereas the rearrangement of Schiff base intermediate to Amadori product takes number of days. An Amadori product undergoes subsequent oxidative modifications (glycoxidations) that are induced by reactive oxygen species (ROS) and reactive nitrogen species (RNS). The end result of these complex series of reactions is the formation of AGEs. Extent of glycation of vital biochemicals is dependent on the degree and duration

of hyperglycemia *in vivo*. Considerable effort has been focused on the discovery of new inhibitors of glycation because of their therapeutic potential ³.

The antiglycating agents may react with carbonyl group of reducing sugars, Amadori products and 3-deoxyglucosones to prevent AGEPs formation. Certain agents have been established that can cleave AGEP cross-links and perhaps open the possibility of moving back the steady process of diabetic complications ⁴. Antioxidants may offer protection against glycated free radicals, while chelators are helpful in removing the transition metals to prevent glycoxidations (auto oxidation) of glucose and Amadori products ^{5,6}.

Compounds with both antiglycation and antioxidant properties may offer therapeutic potential. It has been found that aged garlic extract (AGE) inhibit the formation of AGEPs *in vitro* and it also prevents the formation of glycation-derived free radicals.S-Allylcysteine is a key component of aged garlic extract that acts as a potent antioxidant and may inhibit the AGEPs formation? Aminoguanidine (inhibitor of AGEP formation) was found to prevent retinopathy in diabetic animals and has been withdrawn from phase III clinical trials due to its toxicity, hence can serve as a prototype for novel molecules that are being synthesized and evaluated *in vitro*^{8, 9}.

Quinazoline and its derivatives constitute an interesting molecule among the most important classes of an hetero aromatic bicyclic compounds with wide-ranging biological and pharmacological activities such analgesic¹⁰, anticancer¹¹, anticonvulsant¹², anti-HIV¹³, antimicrobial¹⁴, antioxidant¹⁵, antitubercular¹⁶, antiviral¹⁷, anti-inflammatory ¹⁸ and etc.

1,2,3-Triazoles are an important class of heterocycles due to their wide range of applications as synthetic intermediates and pharmaceuticals¹⁹. Several therapeutically interesting 1,2,3-triazoles have been reported, including anti-HIV agents^{20,21}, antimicrobial

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Scheme 1. Reagents and conditions: (a) aryl aldehyde, NaOH, EtOH, Strring<10 °C, 2-4h; (b) Urea, KOH, EtOH, reflux, 4-6h; (c) semicarbazide HCl/thiosemicarbazide HCl, KOH, EtOH, reflux, 4-5h.

compounds²², β 3-selective adrenergic receptor agonists²³, kinase inhibitors²⁴, ²⁵, aceylcholinesterase inhibitors²⁶, carbonic anhydrase II inhibitors²⁷ and etc. Along with the diverse biological activities of quinazoline and its derivatives, quinazoline fused with other heterocycles play an essential role in several biological processes and have a considerable chemical and pharmacological importance.

Drug discovery is considerably directed towards identification of new and safe synthetic antiglycating agents and is an important area of pharmaceutical research for the treatment of late diabetic complications²⁸⁻³¹. Since the availability of an ideal antiglycating agent is less, it instiates the research on molecules for their antiglycating activity⁶. The main part of our research has been devoted to synthetic methods leading to new ring systems, containing the triazolo hexahydroquinazolines nucleus, as a pharmacophore moiety for potential drugs. Hence, *in vitro* antiglycation and antioxidant activities of some 5-aryl-triazolo [4,3-a] hexahydroquinazolines was considered for present investigation.

2. Results and Discussion

2.1. Chemistry

The final compounds 4a-4e & 5a-5e were synthesized following the synthetic route outlined in **Scheme 1**. All the 2-arylidenecyclohexanone (2a-e) were prepared based on Claisen-Schmidt condensation between the aromatic aldehydes and cyclohexanone. Cyclization of 2a-e with urea in ethanolic KOH provided 3a-e. Reaction of 3a-e with semicarbazide HCl and thiosemicarbazide HCl in ethanolic KOH offered 4a-e and 5a-e respectively and were characterized by their IR, 1H-NMR and ES-MS spectral and elemental analysis data. CHNS microanalysis revealed that variation in experimental values compared with calculated values is within ±0.4%. All the 5-(4-aryl)-4,5,6,7,8,9-hexahydro-[1,2,4]triazolo[4,3-a]quinazolin-1-ol (4a-4e) derivatives showed a characteristic O-H stretch (between 3329-3345 cm⁻¹), NH stretch (between 3123-3201 cm⁻¹) and C=N stretch (between 1637-1658 cm⁻¹). In ¹H-NMR spectrum displayed a characteristic peak for triazolo hydroxy proton (-OH) between δ7.63-8.10 ppm as singlet, hexahydroquinazoline NH proton between δ 5.50-5.72 ppm as singlet, CH proton between δ 4.60-4.81 ppm as singlet and cyclohexyl-CH₂ protons displayed a triplet of multiplet between δ 1.50-3.38 5-(4-aryl)-4,5,6,7,8,9-hexahydroppm. All the [1,2,4]triazolo[*4,3-a*]quinazolin-1-thiol (5a-5e) derivatives showed a characteristic NH stretch (between $3145-3234~\text{cm}^{-1}$), S-H stretch (between 2532-2554 cm⁻¹) and C=N stretch (between 1555-1642 cm⁻¹). In ¹H-NMR spectrum displayed a characteristic peak for triazolo thiol proton (-SH) between δ7.96-8.14

ppm as singlet, CH proton between δ 4.37- 4.70 ppm as singlet, hexahydroquinazoline NH proton between δ 3.90-4.25 ppm as singlet and cyclohexyl-CH₂ protons displayed a triplet of multiplet between δ 1.24-2.80 ppm. The EI-MS spectra of all the compounds **(4a-4e** & **5a-5e)** displayed (M+1)+ peak. The structure and physical characterization of compounds **4a-e** and **5a-e** is presented in **Table 1**.

Table 1. Physical Properties of 4a-e & 5a-e

Code	R	X	MF	MW	MP(°C)
4a	phenyl	0	$C_{15}H_{16}N_4O$	268	220-222
4b	p-OCH₃ phenyl	0	$C_{16}H_{18}N_4O_2\\$	298	186-188
4c	p-Cl phenyl	0	$C_{15}H_{15}ClN_4O$	302	196-198
4d	furan-2-yl	0	$C_{13}H_{14}N_4O_2\\$	258	230-232
4e	p-NO ₂ phenyl	0	$C_{15}H_{15}N_5O_3\\$	313	215-217
5a	phenyl	S	$C_{15}H_{16}N_4S$	284	184-186
5b	p-OCH₃ phenyl	S	$C_{16}H_{18}N_4O_S\\$	314	210-214
5c	p-Cl phenyl	S	$C_{15}H_{15}ClN_4S$	318	164-168
5d	furan-2-yl	S	$C_{13}H_{14}N_4OS$	274	221-223
5e	p-NO ₂ phenyl	S	$C_{15}H_{15}N_5O_2S\\$	329	198-200

2.2. Anti glycating activity

All the synthesized 4a-4e & 5a-5e were randomly evaluated for their in vitro antiglycating activity. The compounds exhibited a varying degree of antiglycating activity, when compared to standards rutin (antiglycating agent; $IC_{50} = 41.9 \mu M$) and glibenclamide agent; 0.002 μM) (Table (antidiabetic Comparatively, all the synthesized compounds are potent than rutin in the range between 0.096 and 0.300 μM. The phenyl ring substituted with electron donating group have shown more potency than electron withdrawing substituent i.e. 4b (p-OCH₃ phenyl) and 5c (p-Cl phenyl), also excellent antiglycation activities, when compared to standard rutin, but less potent than glibenclamide (IC₅₀ = $0.002 \mu M$). Additionally, compounds **4e** (p-NO₂ phenyl) and **5b** (p-OCH₃) initially showed less than 50 % activity and thus were not further evaluated for their IC_{50} . The thiol derivatives are more potent than the corresponding hydroxyl derivatives, as **5a** ($IC_{50} = 0.117 \mu M$) is more potent than **4a** ($IC_{50} = 0.191 \mu M$) and **5c** ($IC_{50} = 0.096 \mu M$) is more potent than 4c (IC₅₀ =0.300 μ M). Compound 4b, 4d, 5a, **5c,** and **5e**have shown more potent anti-glycating activity than all other derivatives. Among these compounds 4b and 5c are more potent. Within this small set of active molecules, a concrete SAR discussion may not be feasible. But few guiding principles can be derived to proceed further in designing this class of molecules as antiglycating agents. The IC50 values of anti-glycating activity given in Table 2.

Table 2. IC_{50} values for antiglycating activity of 4a-e & 5a-e

Code	R	X	IC50 (μM)
4a	phenyl	0	0.191
4b	p-OCH3 phenyl	0	0.104
4c	p-Cl phenyl	0	0.300
4d	furan-2-yl	0	0.134
4e	p-NO2 phenyl	0	ND
5a	phenyl	S	0.117
5b	p-OCH3 phenyl	S	ND
5c	p-Cl phenyl	S	0.096
5d	furan-2-yl	S	0.192
5e	p-NO2 phenyl	S	0.102
RTN	-	-	41.9
GBD	-	-	0.002

ND: Not Determined: RTN-Rutin: GBD-Glibenclamide

Antioxidant activity:

All the synthesized compounds **(4a-4e& 5a-5e)** were tested for antioxidant activity by DPPH assay. These compoundsexhibited potent to moderate antioxidant activity in the present investigation. In the series, almost all the test compounds were found to be potent than BHT (Butylated Hydroxytoluene) except $\mathbf{5e}$ (p-NO₂ phenyl). Among all,the hydroxyl derivatives ($\mathbf{4a-4e}$) are more potent than the corresponding thiols ($\mathbf{5a-5e}$). Molecules $\mathbf{4c}$ (p-Cl phenyl) and $\mathbf{4b}$ (p-OCH₃) were the most potent compounds. The IC₅₀ values were shown in **Table 3**.

Table 3. IC₅₀ values for antioxidant activity of 4a-e & 5a-e

Code	R	X	IC ₅₀ (μM)
4a	phenyl	0	0.213
4b	p-OCH₃ phenyl	0	0.122
4c	p-Cl phenyl	0	0.093
4d	furan-2-yl	0	0.237
4e	p-NO ₂ phenyl	0	0.122
5a	phenyl	S	0.137
5b	p-OCH₃ phenyl	S	0.226
5c	p-Cl phenyl	S	0.269
5d	furan-2-yl	S	0.216
5e	p-NO ₂ phenyl	S	0.589
BHT	-		0.510

BHT-Butylated hydroxy toluene

3. Experimental

Materials and methods: All the reagents and chemicals were obtained commercially from Merck, Spectrochem or CDH, India. Purified solvents of reagent grade were purified, dried by standard procedure. Monitoring of reactions and homogeneity of the intermediate and final compounds were carried out by thin-layer chromatography on silica gel plates and visualized in either iodine or UV chambers. Melting points were determined using Thermonik Melting Point Apparatus (Campbell electronics, India) by open capillary method and are uncorrected. Infrared (IR) spectra were taken on a FT-IR Spectrophotometer IR-Prestige 21 (Shimatzu Corporation, Japan) from 4000-400 cm-1 using KBr discs. ¹H-NMR spectra were recorded at 400 MHz in DMSO-d₆ using a BrukerAvance 400 instrument (Bruker Instruments Inc., USA). Chemical shifts were measured at δ units (ppm) relative to Tetramethylsilane (TMS). Fast-atom bombardment (FAB) mass spectra were recorded on a Jeol SX 102/DA-6000 mass spectrometer (Jeol Ltd. Akishima, Tokyo, Japan) using argon/xenon (6kV, 10mA) as FAB gas, m-nitrobenzyl alcohol as matrix, and 10 kV as accelerating voltage at room temperature. Elemental analysis (%C, H, N) was performed on a Vario EL III Elemental Analyser (Elementar, Germany). The 5-aryl-triazolo [4,3-a]hexahydroquinazolines were synthesized as per **Scheme 1** and characterized by elemental analysis, IR, $^1\text{H-NMR}$ and EI-MS. In the elemental analysis, the observed values were within $\pm 0.4\%$ of the calculated values.

3.1. Chemistry

3.1.1. General procedure for synthesis of 2-arylidenecyclohexanone (2a-2e)

Cyclohexanone (0.01 mole), ethanolic NaOH (20 %; 20ml) were taken in a round bottom flask and drop wise addition of respective benzaldehyde (0.01 mole) with continuous stirring for 2 h. Further stirred was continued for 1 hr and mixture was kept in the refrigerator for overnight. The obtained yellowish precipitate was collected by filtration and crude product was recrystallized with chloroform.

3.1.2. General procedure for synthesis of 4-aryl-3,4,5,6,7,8-hexahydroquinazolin-2(1H)-one (3a-3e):

The 2-arylidenecyclohexanone (2, 0.01 mol), urea (0.02 mol) were taken in ethanolic KOH (20 %; 20 ml) and heated to reflux for 4-6 h. Completion of reaction was monitored by TLC and excess of solvent was removed *in vacuo*. To the residue, cold water (25 ml) was added and neutralized with dil. HCl, then kept in refrigerator icechest for overnight. Collected the crystalline product of 3 by vacuum filtration and further purified by recrystallization with chloroform.

3.1.3. General procedure for synthesis of 5-(4-aryl)-4,5,6,7,8,9-hexahydro-[1,2,4]triazolo[4,3-a]quinazolin-1-ol (4a-4e):

A mixture of 4-aryl-3,4,5,6,7,8-hexahydroquinazolin-2(1H)-ones (3, 0.005 M) and semicarbazide HCl (0.01 M) were dissolved in ethanolic KOH (10 %; 20 ml) in a round bottom flask. The reaction mixture was heated to reflux for 4-5 hrs and progress of reaction was monitored by TLC. After completion of reaction, the mixture was cooled to room temperature then poured into beaker containing ice cold water and neutralized with the addition dil HCl. Precipitate obtained was filtered and dried. Spectroscopic and elemental analysis data of the synthesized compounds 4a-4e given below:

3.1.4. General procedure for synthesis of 5-(4-aryl)-4,5,6,7,8,9-hexahydro-[1,2,4]triazolo[4,3-a]-quinazolin-1-thiol (5a-5e):

A mixture of 4-aryl-3,4,5,6,7,8-hexahydroquinazolin-2(1H)-ones(3, 0.005 M) and thiosemicarbazide HCl (0.01 M) were dissolved in ethanolic KOH (10 %; 20 ml) in a round bottom flask. The reaction mixture was heated to reflux for 4-5 hrs and progress of reaction was monitored by TLC. After completion of reaction, the mixture was cooled to room temperature then poured into beaker containing ice cold water and neutralized with the addition of dil HCl. Precipitate obtained was filtered and dried. Spectroscopic and elemental analysis data of the synthesized compounds 5a-5e given below:

5-phenyl-4,5,6,7,8,9-hexahydro[1,2,4]triazolo[4,3-a]quinazo lin-1-ol **(4a)**: IR (KBr, cm⁻¹): 3329 (O-H stretch), 3123 (NH stretch), 1652 (C=N stretch); ¹H-NMR (DMSO-d6, δ ppm): 7.90 (s, 1H, OH), 6.60-7.40 (m,

5H, Ar.H), 4.78 (s, 1H, 1NH), 3.75 (s, 1H, CH), 2.25-2.38 (m, 4H, cyclohexyl-CH₂), 1.88-1.96 (t, 4H, cyclohexyl-CH₂); Anal. Calcd. for $C_{15}H_{16}N_4O$ (268.31): C, 67.15; H, 6.01; N, 20.88%. Found: C, 67.28; H, 5.90; N, 20.92%; MS m/z: 269 (M+1)+

5-(4-methoxyphenyl)-4,5,6,7,8,9-hexahydro- [1,2,4] triazolo [4,3-a]quinazolin-1-ol (4b): IR (KBr, cm $^{-1}$): 3334 (O-H stretch), 3128 (NH stretch), 1644 (C=N stretch); 1 H-NMR (DMSO-d $_{6}$, δ ppm): 8.10 (s, 1H, OH), 6.41-7.11 (m, 4H, Ar.H), 5.60 (s, 3H, OCH $_{3}$), 4.81 (s, 1H, 1NH), 3.90 (s, 1H, CH), 1.50-3.20 (m, 8H, cyclohexyl-CH $_{2}$); Anal. Calcd. for $C_{16}H_{18}N_{4}O_{2}$ (298.34): C, 64.41; H, 6.08; N, 18.78%. Found: C, 64.58; H, 6.09; N, 18.84%; MS m/z: 299 (M+1)+

5-(4-chlorophenyl)-4,5,6,7,8,9-

hexahydro[1,2,4]triazolo[4,3 -a]quinazolin-1-ol (4c): IR (KBr, cm $^{-1}$): 3330 (O-H stretch), 3201 (NH stretch), 1640 (C=N stretch); 1 H-NMR (DMSO-d6, δ ppm): 7.90 (s, 1H, OH), 7.43 (dd, 2H, Ar.H), 7.35 (dd, 2H, Ar.H), 4.67 (s, 1H, 1NH), 3.80 (s, 1H, CH), 2.25-2.38 (m, 4H, cyclohexyl-CH₂), 1.88-1.96 (t, 4H, cyclohexyl-CH₂); Anal. Calcd.for C₁₅H₁₅ClN₄O (302): C, 59.51; H, 4.99; N, 18.51%. Found: C, 59.72; H, 5.00; N, 18.56%; MS m/z: 303 (M+1) $^{+}$

5-(furan-2-yl)-4,5,6,7,8,9-hexahydro[1,2,4]triazolo[4,3-a]quinazolin-1-ol (4d): IR (KBr, cm⁻¹): 3338 (O-H stretch), 3143 (NH stretch), 1658 (C=N stretch); 1 H-NMR (DMSO-d6, δ ppm): 8.10 (s, 1H, OH), 7.80 (d, 1H, Ar.H), 6.52-7.12 (m, 2H, ArH), 4.76 (s, 1H, 1NH), 3.75 (s, 1H, CH), 1.64-3.38 (m, 8H, cyclohexyl-CH₂); Anal. Calcd. for C₁₃H₁₄N₄O₂ (258.27): C, 60.45; H, 5.46; N, 21.69%. Found: C, 60.64; H, 5.45; N, 21.72%; MS m/z: 259 (M+1)+

5-(4-nitrophenyl)-4,5,6,7,8,9-

hexahydro[1,2,4]triazolo[4,3-a]quinazolin-1-ol (4e): IR (KBr, cm⁻¹): 3345 (O-H stretch), 3168 (NH stretch), 1637 (C=N stretch); 1 H-NMR (DMSO-d₆, δ ppm): 7.63 (s, 1H, OH), 7.36-7.43 (m, 4H, ArH), 4.60 (s, 1H, 1NH), 3.70 (s, 1H, CH), 2.58-2.64 (m, 4H, cyclohexyl-CH₂), 1.86-1.89 (t, 4H, cyclohexyl-CH₂); Anal. Calcd.for C₁₅H₁₅N₅O₃ (313.31): C, 57.50; H, 4.83; N, 22.35%. Found: C, 57.48; H, 4.82; N, 22.42%; MS m/z: 314 (M+1) $^{+}$

5-phenyl-4,5,6,7,8,9-hexahydro[1,2,4]triazolo[4,3-a]quinazoline-1-thiol (5a): IR (KBr, cm⁻¹): 3221 (NH stretch), 2547 (S-H stretch), 1575 (C=N stretch); ¹H-NMR (DMSO-d6, δ ppm): 8.02 (s, 1H, SH), 6.82-7.80 (m, 5H, Ar-H), 4.68 (s, 1H, 1NH), 3.75 (s, 1H, CH), 2.00-2.60 (m, 4H, cyclohexyl-CH₂), 1.64-1.84 (t, 4H, cyclohexyl-CH₂); Anal. Calcd.for C₁₅H₁₆N₄S (284.37): C, 63.35; H, 5.67; N, 19.70%. Found: C, 63.42; H, 5.68; N, 19.74%; MS m/z: 285 (M+1)+

5-(4-methoxyphenyl)-4,5,6,7,8,9-

hexahydro[1,2,4]triazolo[4,3-a]quinazoline-1-thiol (5b): IR (KBr, cm-¹): 3234 (NH stretch), 2554 (S-H stretch), 1560 (C=N stretch); 1 H-NMR (DMSO-d6, δ ppm): 7.96 (s, 1H, SH), 6.76-7.82 (m, 4H, Ar.H), 5.65 (s, 3H, OCH₃), 4.37(s, 1H, 1NH), 3.80 (s, 1H, CH), 2.00-2.60 (m, 4H, cyclohexyl-CH₂), 1.64-1.84 (t, 4H, cyclohexyl-CH₂); Anal. Calcd.for C₁₆H₁₈N₄OS (314.41): C, 61.12; H, 5.77; N, 17.82%. Found: C, 61.26; H, 5.78; N, 17.76%; MS m/z: 315 (M+1)+

5-(4-chlorophenyl)-4,5,6,7,8,9-

hexahydro[1,2,4]triazolo[4,3-a]quinazoline-1-thiol (5c): IR (KBr, cm⁻¹): 3216 (NH stretch), 2543 (S-H stretch),

1555 (C=N stretch); ¹H-NMR (DMSO-d6, δ ppm): 8.12 (s, 1H, SH), 7.12-7.78 (m, 4H, Ar.H), 4.41(s, 1H, 1NH), 3.90 (s, 1H, CH), 1.56-2.80 (m, 8H, cyclohexyl-CH₂); Anal. Calcd.forC₁₅H₁₅ClN₄S (318): C, 56.51; H, 4.74; N, 17.57%. Found: C, 56.48; H, 4.75; N, 17.54%; MS *m/z*: 319 (M+1)+

5-(furan-2-yl)-4,5,6,7,8,9-hexahydro[1,2,4]triazolo[4,3-a]quinazoline-1-thiol (5d): IR (KBr, cm⁻¹): 3145 (NH stretch), 2532 (S-H stretch), 1642 (C=N stretch); ¹H-NMR (DMSO-d6, δ ppm): 8.14 (s, 1H, SH), 7.80 (d, 1H, Ar.H), 6.58-7.10 (m, 2H, Ar.H), 4.70 (s, 1H, 1NH), 3.90 (s, 1H, CH), 1.42-1.64 (m, 4H, cyclohexyl-CH₂), 1.24-1.36 (m, 4H, cyclohexyl-CH₂); Anal. Calcd.for C₁₃H₁₄N₄OS (274.34): C, 56.91; H, 5.14; N, 20.42%. Found: C, 57.10; H, 5.15; N, 20.44%; MS m/z: 275 (M+1)+

5-(4-nitrophenyl)-4,5,6,7,8,9-

hexahydro[1,2,4]triazolo[4,3-a]quinazoline-1-thiol (5e): IR (KBr, cm $^{-1}$): 3226 (NH stretch), 2550 (S-H stretch), 1576 (C=N stretch); 1 H-NMR (DMSO-d6, δ ppm): 8.09 (s, 1H, SH), 7.26-7.89 (m, 4H, Ar.H), 4.40 (s, 1H, 1NH), 4.20 (s, 1H, CH), 1.6-1.9 (t, 4H, 2CH $_2$) 1.20-1.40 (m, 4H, 2CH $_2$); Anal. Calcd.for C₁₅H₁₅N₅O₂S (329.37): C, 54.70; H, 4.59; N, 21.26%. Found: C, 54.88; H, 4.60; N, 21.30%; MS m/z: 330 (M+1) $^{+}$

3.2. Anti-glycating activity

In all experiments, the final reaction volume was $60 \mu L$ and the reactions were performed in 1.5 ml microcentrifuge tubes. Each tube contains, BSA (20 µL; 10 mg/mL), glucose anhydrous (20 μL; 50 mg/mL) and test sample (20 μ L) then final volume maede upto 60 μ L with DMSO. Glycated control contains 20 μL BSA, 20 μL glucose and 20 μL sodium phosphate buffer, while blank control contains 20 µL BSA and 40µL sodium phosphate buffer. Glibenclamide was used as the standard. Reaction mixture was incubated at 37 °C for seven days. After incubation, 6 µL (100%) of TCA was added in each tube and centrifuged (15,000 rpm) for four minutes at 4 °C. After centrifugation, the obtained pellets were rewashed with 60 µL (10%) of TCA. The supernatant containing glucose, inhibitor and interfering substance was removed and pellet containing AGE-BSA was dissolved in 60 µL PBS. Assessment of fluorescence spectrum (ex. 370 nm) and change in fluorescence intensity (ex. 370 nm to em. 440 nm) based on AGEs were monitored by using a spectrofluorimeter (RF-1500). The % Inhibition was calculated through the following formula³² and IC₅₀ values were given in **Table**

 $\%\ Inhibition = 1 - \frac{Fluorescence\ of\ sample}{Fluorescence\ of\ glycalated\ sample} x 100$

3.3. Antioxidant activity

The antioxidant activity of the synthesized compounds (3a-3e & 4a-4e) and the standard was assessed on the basis of the radical scavenging effect of 1, 1-diphenyl-2-picryl-hydrazyl (DPPH)-free radical activity by modified method 33 . The solutions of the test were prepared in methanol. BHT was used as standard. 0.135 mM DPPH was prepared in methanol and 1 ml of prepared solution of DPPH mixed with 3 ml of methanolic solution of test solutions (20-100 $\mu g/ml$) and BHT (Butylated Hydroxytoluene) solution separately. These mixtures were vortexed thoroughly and kept in dark at room temperature for 30 min. the absorbance was measured at 517 nm using Schimadzu UV-1800

Spectrophotometer. Methanol (1 ml) with DPPH solution (1 ml) was used as blank. The optical density was recorded and % inhibition was calculated using the formula given below³⁴:

% Inhibition of DPPH activity =
$$\frac{[A_0 - A_s]}{A_0} \times 100$$

Where A_0 is the absorbance of control and A_s is the absorbance of sample.

4. Conclusion

Advanced glycation end products are the main cause for diabetic complications. Aminoguanidine is the prototype molecule for use in the treatment of these complications like retinopathy. In the present study, triazolo- hexahydroquinazolines were synthesized as isosteric modification of the aminoguanidines and evaluated them for antiglycating and antioxidant activities. The molecules **4b** and **5c** were found to have potent antiglycating activity than rutin and less potent than glibenclamide due to the presence of electron donating groups substituted on the benzene ring. The molecules **4b** and **4c** were found to have potent antioxidant activity due to the presence of hydroxyl groups. The results obtained may contribute to the development of an ideal antiglycating agents.

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References

- Brownlee, M. Lilly Lecture 1993. Glycation and diabetic complications. Diabetes 1994, 43, 836-41.
- Peppa, M.; Uribarri, J.; Vlassara, H. Glucose, Advanced Glycation End Products, and Diabetes Complications: What Is New and What Works. Clin. Diabetes 2003, 21, 186-187.
- Monnier, V. M. Intervention against the Maillard reaction in vivo. Arch Biochem Biophys 2003, 419, 1.15
- Vasan, S.; Foiles, P.; Founds, H. Therapeutic potential of breakers of advanced glycation end productprotein crosslinks. Arch Biochem Biophys 2003, 419, 89-96.
- Hunt, J. V.; Bottoms, M. A.; Mitchinson, M. J. Oxidative alterations in the experimental glycation model of diabetes mellitus are due to protein-glucose adduct oxidation. Some fundamental differences in proposed mechanisms of glucose oxidation and oxidant production. Biochem J 1993, 291 (Pt 2), 529-35.
- Ahmed, N. Advanced glycation endproducts--role in pathology of diabetic complications. Diabetes Res Clin Pract 2005, 67, 3-21.
- Ahmad, M. S.; Ahmed, N. Antiglycation properties of aged garlic extract: possible role in prevention of diabetic complications. J Nutr 2006, 136, 796S-799S.
- Gugliucci, A. Glycation as the glucose link to diabetic complications. J Am Osteopath Assoc 2000, 100, 621-634.
- 9. Singh, R.; Barden, A.; Mori, T.; Beilin, L. Advanced glycation end-products: a review. Diabetologia 2001, 44, 129-146.
- Alagarsamy, V.; Muthukumar, V.; Pavalarani, N.; Vasanthanathan, P.; Revathi, R.; Synthesis, analgesic and anti-inflammatory activities of some novel 2,3-

- disubstituted quinazolin-4(3H)-ones. Biol Pharm Bull 2003, 26, 557-559.
- 11. Shetha, A.; Wijdan, I. A. Synthesis and characterization of new quinazoline-4(3H)-one Schiff bases. J Chem Pharm Res 2013, 5, 42–45.
- 12. Patel, N. B.; Patel, V. N.; Patel, H. R.; Shaikh, F. M.; Patel, J. C. Synthesis and microbial studies of (4-oxothiazolidinyl) sulfonamides bearing quinazolin-4(3H)ones. Acta Pol Pharm 2010, 67, 267–275.
- 13. Pati, B.; Banerjee, S. Quinazolines: an illustrated review. J Adv Pharm Edu & Res 2013, 3, 136–51.
- Rudrapal, M.; De, B. Chemistry and biological importance of heterocyclic Schiff's bases. Int Res J Pure & App Chem 2013, 3, 232–249.
- Vagdevi, H. M.; Lokesh, M. R.; Gowdarshivannanavar, B. C. Synthesis and antioxidant activity of 3-substituted Schiff bases of quinazoline-2,4-diones. Int J Chem Tech Res 2012, 4, 1527–1533.
- Abid, O. H.; Ahmed, A. H. Synthesis and characterization of novel quinazoline derivatives via reaction of isatoic anhydride with schiff's base. Int J App Nat Sci 2013, 2, 11-20.
- 17. Krishnan, S. K.; Ganguly, S.; Veerasamy, R.; Jan, B. Synthesis, antiviral and cytotoxic investigation of 2-phenyl-3-substituted quinazolin-4(3H)-ones. Eur Rev Med Pharmacol Sci 2011, 15, 673–681.
- 18. Saravanan, G.; Pannerselvam, P.; Prakash, C. R. Synthesis, analgesic and anti-inflammatory screening of novel Schiff bases of 3-amino-2-methyl quinazoline 4-(3H)-one. Der Pharmacia Lettre 2010, 2, 216–226.
- Fan, W. Q.; Katritzky, A. R. 1,2,3-Triazoles. In Comprehensive Heterocycle Chemistry II; Katritzky, A.R.; Rees, C. W.; Scriven, E.F.V.; Eds.; Pergamon Press: New York, NY, USA, 1996; 4, 1–126.
- Biorn, A. C.; Cocklin, S.; Madani, N.; Si, Z.; Ivanovic, T.; Samanen, J.; Ryk, D. I. V.; Pantophlet, R.; Burton, D. R.; Freire, E.; Sodroski, J.; Chaiken, I. M. Mode of action for linear peptide inhibitors of HIV-1 gp120 interactions. Biochemistry 2004, 43, 1928–1938.
- Whiting, M.; Muldoon, J.; Lin, Y. C.; Silverman, S. M.; Lindstrom, W.; Olson, A. J.; Kolb, H. C.; Finn, M. G.; Sharpless, B. K.; Elder, J. H.; Fokin, V. V. Inhibitors of HIV-1 protease by using in situ click chemistry. Angew. Chem. Int. Ed. 2006, 45, 1435–1439.
- Wang, Z. J.; Gao, Y.; Hou, Y. L.; Zhang, C.; Yu, S. J.; Bian, Q.; Li, Z. M.; Zhao, W. G. Design, synthesis, and fungicidal evaluation of a series of novel 5-methyl-1H-1,2,3-trizole-4-carboxyl amide and ester analogues. Eur. J. Med. Chem. 2014, 86, 87–94.
- Brockunier, L. L.; Parmee, E. R.; Ok, H. O.; Candelore, M. R.; Cascieri, M. A.; Colwell, L. F.; Eng, L.; Feeney, W. P.; Forrest, M. J.; Hom, G. J.; MacIntyre, D. E.; Tota, L.; Wyvratt, M. J.; Fisher, M. H.; Weber, A. E. Human beta3-adrenergic receptor agonists containing 1,2,3-triazole substituted benzenesulfonamides. Bioorg. Med. Chem. Lett. 2000, 10, 2111–2114.
- 24. Pande, V.; Ramos, M. J. Structural basis for the GSK-3beta binding affinity and selectivity against CDK-2 of 1-(4-aminofurazan-3yl)-5-dialkylaminomethyl-1H-[1,2,3]triazole-4-carboxylic acid derivatives. Bioorg. Med. Chem. Lett. 2005, 15, 5129–5135.
- Olesen, P. H.; Sørensen, A. R.; Ursö, B.; Kurtzhals, P.; Bowler, A. N.; Ehrbar, U.; Hansen, B. F. Synthesis and in vitro characterization of 1-(4-Aminofurazan-3yl)-5-dialkylamino-methyl-1H- [1,2,3]triazole-4carboxylic acid derivatives. A new class of selective GSK-3 inhibitors. J. Med. Chem. 2003, 46, 3333-3341
- Krasinski, A.; Radic, Z.; Manetsch, R.; Raushel, J.; Taylor, P.; Sharpless, B. K.; Kolb, H. C. In situ selection of lead compounds by click chemistry: Target-guided optimization of aceylcholinesterase inhibitors. J. Am. Chem. Soc. 2005, 127, 6686–6692.
- 27. Mocharla, V. P.; Colasson, B.; Lee, L. V.; Roeper, S.; Sharpless, B. K.; Wong, C. H.; Kolb, H. C. In situ click

- chemistry: Enzyme-generated inhibitors of carbonic anhydrase II. Angew. Chem. Int. Ed. 2005, 44, 116–120
- Degenhardt, T. P.; Alderson, N. L.; Arrington, D. D.;
 Beattie, R. J.; Basgen, J. M.; Steffes, M. W.; Thorpe, S.
 R.; Baynes, J. W. Pyridoxamine inhibits early renal disease and dyslipidemia in the streptozotocin-diabetic rat. Kidney Int 2002, 61, 939-950.
- Forbes, J. M.; Soulis, T.; Thallas, V.; Panagiotopoulos, S.; Long, D. M.; Vasan, S.; Wagle, D.; Jerums, G.; Cooper, M. E. Renoprotective effects of a novel inhibitor of advanced glycation. Diabetologia 2001, 44, 108-14.
- Lehman, T. D.; Ortwerth, B. J. Inhibitors of advanced glycation end product-associated protein crosslinking. Biochim Biophys Acta 2001, 1535, 110-119.
- Price, D. L.; Rhett, P. M.; Thorpe, S. R.; Baynes, J. W. Chelating activity of advanced glycation end-product inhibitors. J Biol Chem 2001, 276, 48967-48972.
- 32. Nakagawa, T.; Yokozawa, T.; Terasawa, K.; Shu, S.; Juneja, L. R. Protective activity of green tea against free radical- and glucose-mediated protein damage. J Agric Food Chem 2002, 50, 2418-2422.
- Braca, A.; Sortino, C.; Politi, M.; Morelli, I.; Mendez, J. Antioxidant activity of flavonoids from *Licania licaniaeflora*. J Ethnopharmacol 2002, 79, 379-381.
- Bors, W.; Saran, M.; Elstner, E. Screening for plant antioxidants. Modern Methods of Plant Analysis-Plant Toxin Analysis-New Series 1992, 13, 277-295.